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54 Peptide fragments and analogs of thrombospondin.

57 Compounds and compositions comprising fragments and synthetic analogs of human thrombospondin are provided together with methods for their use as thrombospondin-like agents.

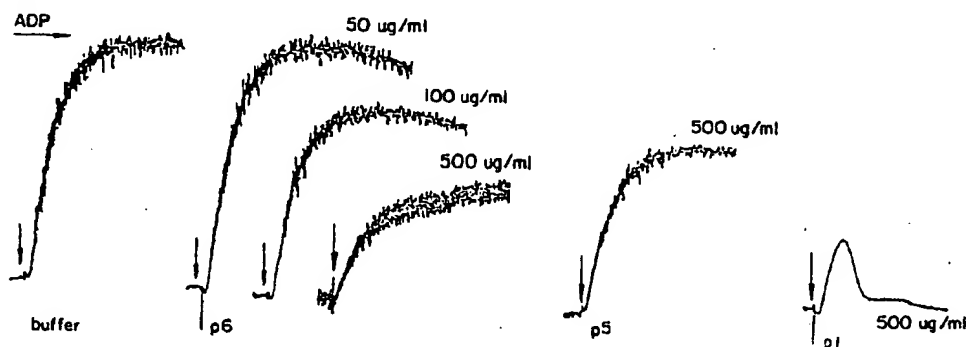


FIG. 1

Technical Field

The present invention relates generally to peptide fragments and synthetic analogs of thrombospondin which retain thrombospondin-like activity. The peptides find use as agents in anti-tumor development, wound healing, atherosclerosis, antithrombotics, thrombolytics, angiogenesis, complement activation, cell attachment, diagnostic reagents and other related areas.

Background

Thrombospondin (also known as thrombin sensitive protein or TSP) is a large molecular weight 180 kD glycoprotein composed of three identical disulfide-linked polypeptide chains. Thrombospondin has been purified by a number of procedures including exclusion chromatography (Lawler et al., *J. Biol. Chem.* (1978) 253:8609-16), heparin affinity chromatography (Lawler et al., *Thromb. Res.* (1981) 22:267-269), fibrinogen affinity chromatography (Tuszynski et al., *J. Biol. Chem.* (1985) 260:12240-5), barium chloride precipitation (Alexander et al., *Biochem. J.* (1984) 217:67-71) and anion exchange chromatography with HPLC (Ciezarolin et al., *J. Chromatog.* (1984) 296:249-56).

The complete amino acid sequence of TSP has been deduced from DNA clones prepared by various groups including Lawler et al., *J. Cell Biol.* (1986) 103:1635-48; Kobayashi et al., *Biochemistry* (1986) 25:8418-25; Dixit et al., *Proc. Natl. Acad. Sci.* (1986) 83:5449-53; and Hennessy et al., *J. Cell Biol.* (1989) 108:729-36.

Kobayashi et al. (*supra*), Robson et al. (*Nature* (1988) 335:79-82) and Goundis et al. (*Nature* (1988) 335:82-85) compared the sequences of thrombospondin, thrombospondin-related anonymous protein, properdin, the terminal complement components, and circumsporozoite proteins and recognized a significant homology based around the consensus sequence (SEQ ID No. 1) W-S-P-C-S-V-T-C-G. While the sequence homology was recognized, there was no known use for such sequence.

The native thrombospondin molecule has been shown to potentiate tumor cell metastasis (Tuszynski et al., *Cancer Research* (1987) 47:4130-4133). The mechanisms by which the thrombospondin potentiation occurs are not presently well understood.

Thrombospondin and/or TSP-like proteins have been reported to have effects in the inhibition of angiogenesis (Rastinejad et al., *Cell* (1989) 56:345-355, and Good et al., *J. Cell Biol. Suppl.* (1989) 13B:30). The mechanism by which thrombospondin modulates angiogenesis is not well understood.

The present invention provides thrombospondin fragments and analogs which mimic or inhibit the biological activity of intact thrombospondin which find use in a variety of biological, prophylactic or therapeutic areas.

Summary of the Invention

It has now been found that a class of fragments or synthetic analogs of thrombospondin have a variety of uses. These peptides are capable of inhibiting tumor metastasis in mammals *in vivo*. The peptides are also useful in wound healing, atherosclerosis, thrombotic, and thrombolytic conditions, angiogenesis, and as cell attachment promoters, complement modulators, and diagnostic reagents and in other related areas. Analogs based on the Type I repeat of thrombospondin described by Lawler et al., (*Seminars in Thrombosis & Hemostasis* (1987) 13:245-254), Robson et al., *supra*, and Goundis et al., *supra*, have been shown to have thrombospondin-like activity. Specifically, analogs based around and including at least a portion of the sequence motif of Robson (*supra*) W-S-P-C-S-V-T-C-G have been shown to have thrombospondin-like activity.

The present invention is, therefore, in one aspect directed to polypeptide compounds having thrombospondin-like activity which are identified by the formula:

Z<sub>1</sub>-AA<sub>1</sub>-AA<sub>2</sub>-AA<sub>3</sub>-AA<sub>4</sub>-AA<sub>5</sub>-AA<sub>6</sub>-AA<sub>7</sub>-AA<sub>8</sub>-AA<sub>9</sub>-Z<sub>2</sub>

wherein:

- AA<sub>1</sub> is a neutral/non-polar/large/cyclic amino acid residue;
- AA<sub>2</sub> is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;
- AA<sub>3</sub> is a neutral/nonpolar/large/cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- AA<sub>4</sub> is a neutral/polar/small amino acid residue;
- AA<sub>5</sub> is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;

- AA<sub>6</sub> is a neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic amino acid residue;  
 AA<sub>7</sub> is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;  
 AA<sub>8</sub> is a neutral/polar/small amino acid residue;  
 AA<sub>9</sub> is a neutral/polar/small amino acid residue;  
 5 Z<sub>1</sub> is hydrogen, amino, acetyl or at least one amino acid residue or the desamino form thereof;  
 Z<sub>2</sub> is hydroxyl, carboxyl, non-amino acids such as agmatine, or at least one amino acid residue,  
 including carboxamide or alkylamide forms thereof.

Also provided in accordance with aspects of the invention are pharmaceutical compositions which contain the above-recited polypeptide compounds together with a pharmaceutically acceptable liquid, gel or  
 10 solid carrier. Administration of therapeutically effective doses of these compositions can provide effective enhancement or inhibition of thrombospondin-like activity to animals, particularly vertebrates such as mammalian and avian hosts.

#### Brief Description of the Drawings

15 Figure 1 depicts the in vitro ability of selected peptide compounds of the present invention to inhibit platelet aggregation.

#### Detailed Description of the Invention

20 in accordance with the present invention, a class of fragments and analogs of thrombospondin is provided which is capable of inhibiting or mimicing the activity of thrombospondin in mammals in vivo.

#### A. Definitions

25 "Thrombospondin-like activity" is defined herein as any activity that mimics the known biological activities of thrombospondin. These activities include cell-adhesion promoting activity, cell mitogenic activity, cell chemotactic activities, and hemostatic activities and any activities that derive from these activities such as tumor cell or microbial metastasis activity, platelet aggregating activity, fibrinolytic activity  
 30 and immune modulation.

"Antimetastatic activity" is defined herein as the ability to prevent or greatly reduce the extent or size of tumor cell metastasis, or inhibit or cause regression of primary solid tumors.

"Wound healing activity" is defined herein as the ability to increase the rate at which wounds heal or to improve the results of the healing process (i.e., less scarring, good response to tactile stimulus, etc.)

35 "Atherosclerosis activity" is defined herein as the capacity of thrombospondin to either promote or inhibit atherosclerotic lesion formation. The atherosclerotic lesion is defined as the degenerative accumulation of lipid-containing materials, especially in arterial walls.

"Antithrombotic activity" is defined herein as the ability to either inhibit the aggregation of platelets or to antagonize the formation of a thrombus.

40 "Thrombolytic activity" is defined herein as the ability to disrupt the structure of a thrombus.

"Angiogenesis activity" is defined herein as the ability to inhibit or enhance the formation of blood vessels or lymph vessels.

"Growth factor activity" is defined herein as the ability to inhibit or promote cell proliferation.

45 "Cell adhesion activity" is defined herein as the ability to promote or inhibit the attachment of cells, preferably mammalian cells, to a substrate.

"Complement activity" is defined herein as the ability to activate or block the Complement Cascade Pathway of the immune system.

"Antiviral activity" is defined herein as the ability to prevent or inhibit viral infection by interfering with the ability of the viral particle to bind to cells.

50 The sequence of amino acid residues of the present polypeptide compounds, the core nonapeptide, and preferred embodiments thereof, are defined in terms of amino acids of certain characteristics of particular subclasses.

Amino acid residues can be generally subclassified into four major subclasses as follows:

55 Acidic, i.e., the residue has a negative charge due to loss of H ion at physiological pH and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium.

Basic, i.e., the residue has a positive charge due to association with H ion at physiological pH and the residue is attracted by aqueous solution so as to seek the surface positions in the conformation of a peptide

in which it is contained when the peptide is in aqueous medium.

Neutral/non-polar, i.e., the residues are not charged at physiological pH and the residue is repelled by aqueous solution so as to seek the inner positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium.

- 5 Neutral/polar, i.e., the residues are not charged at physiological pH and the residue is attracted by aqueous solution so as to seek the outer positions in the conformation of a peptide in which it is contained when the peptide is in aqueous medium.

It is understood, of course, that in a statistical collection of individual residue molecules some molecules will be charged, and some not. To fit the definition of charged, a significant percentage (at least 10 approximately 25%) of the individual molecules are charged at physiological pH.

Amino acid residues can be further subclassified as cyclic or non-cyclic, a self-explanatory classification with respect to the side chain substituent groups of the residues, and as small or large. The residue is considered small if it contains a total of three carbon atoms or less. Small residues are, of course, always non-cyclic.

- 15 For the naturally occurring protein amino acids, subclassification according to the foregoing scheme is as follows:

Acidic: Aspartic acid and Glutamic acid;

Basic/non-cyclic: Arginine and Lysine;

Basic/cyclic: Histidine;

- 20 Neutral/polar/small: Glycine, Serine and Cysteine;

Neutral/polar/large/non-cyclic: Threonine, Asparagine and Glutamine;

Neutral/polar/large/cyclic: Tyrosine;

Neutral/non-polar/small: Alanine;

Neutral/non-polar/large/non-cyclic: Valine, Isoleucine, Leucine and Methionine;

- 25 Neutral/non-polar/large/cyclic: Phenylalanine and Tryptophan.

The protein amino acid proline, although within the classification neutral/non-polar/large/cyclic, is not included as an alternative due to its known effects on the secondary conformation of peptide chains.

- Certain commonly encountered non-natural amino acids, such as desamino Tyrosine (des Tyr), agmatine (Agm), n-formyl tryptophan (f-Trp), alpha-aminoisobutyric acid (Aib), and sarcosine (Sar), statine, 30 ornithine (Orn), homolysine, homoserine, homoarginine, norleucine (Nle), norvaline may also be incorporated into the compounds of the invention. Desamino tyrosine is incorporated at the N-terminus. Agmatine and statine are incorporated at the C-terminus. Based on the above definition, n-formyl Trp is neutral/non-polar/large/ cyclic, Sar is neutral/non-polar/small, Aib is neutral/non-polar/non-cyclic, Orn is basic/non-cyclic, homolysine is basic/non-cyclic, homoserine is neutral/polar/small, homoarginine is basic/non-cyclic, nor- 35 leucine is neutral/non-polar/large/non-cyclic, and norvaline is neutral/non-polar/large/non-cyclic.

- The nomenclature used to describe polypeptide compounds of the present invention follows the conventional practice wherein the amino group is presented to the left and the carboxy group to the right of each amino acid residue. In the formulae representing selected specific embodiments of the present invention, the amino- and carboxy-terminal groups, although not specifically shown, will be understood to be 40 in the form they would assume at physiologic pH values, unless otherwise specified. In the amino acid structure formulae, each residue is generally represented by a single letter designation, corresponding to the trivial name of the amino acid, in accordance with the following schedule:

45

50

55

	<u>Amino Acid</u>	<u>One-letter Symbol</u>	<u>Three-Letter Code</u>
5	Alanine	A	Ala
	Arginine	R	Arg
	Asparagine	N	Asn
	Aspartic acid	D	Asp
10	Cysteine	C	Cys
	Glutamine	Q	Gln
	Glutamic acid	E	Glu
15	Glycine	G	Gly
	Histidine	H	His
	Isoleucine	I	Ile
20	Leucine	L	Leu
	Lysine	K	Lys
	Methionine	M	Met
25	Phenylalanine	F	Phe
	Proline	P	Pro
	Serine	S	Ser
30	Threonine	T	Thr
	Tryptophan	W	Trp
	Tyrosine	Y	Tyr
35	Valine	V	Val

In the present application, the L-form of any amino acid residue having an optical isomer is intended unless otherwise expressly indicated, e.g., by the symbol "[D-AA<sub>n</sub>]."

Compounds within the scope of the present invention can be obtained by modifying the disclosed  
 40 formulae in numerous ways, while preserving the activity of the polypeptide compounds thus obtained. For example, while the amino acids of these compounds are normally in the natural L optical isomer form, one or more, usually two or less and preferably one amino acid may be replaced with the optical isomer D form, or a D,L-racemic mixture can be provided in the molecules comprising the polypeptide compound. Additionally, a disulfide linkage may be present or absent in the compounds of the invention, as long as  
 45 activity is maintained.

Amino acid residues contained within the compounds, and particularly at the carboxy- or amino-terminus, can also be modified by methylation, amidation, acetylation or substitution with other chemical groups which can, for example, change the circulating half-life, resistance to proteases and solubility of the compounds without adversely affecting their activity.

50 In addition to the preceding definitions, the following abbreviations have been used throughout in describing the invention:

BCA	bicinchoninic acid
BSA	bovine serum albumin
t-Boc	t-butyloxycarbonyl
55 Bzl	benzyl
°C	degrees centigrade
DCM	dichloromethane
DIEA	diisopropyl ethyl amine

	DMEM	Dulbecco's minimum essential medium
	DMF	dimethyl formamide
	HF	hydrogen fluoride
	HOBT	l-hydroxybenzotriazole
5	HPLC	high performance liquid chromatography
	mBHA	methylbenzhydramine
	μg	microgram
	μl	microliter
	ml	milliliter
10	mM	millimolar
	nm	nanometers
	NMP	N-methylpyrrolidone
	%	percent
	PAM	phenylacetamidomethyl
15	PBS	phosphate buffered saline
	TFA	trifluoroacetic acid

### B. Preferred Embodiments

20 The polypeptide compounds of the invention all contain the core nonapeptide sequence:



wherein:

- 25 AA<sub>1</sub> is a neutral/nonpolar/large/cyclic amino acid residue;
- AA<sub>2</sub> is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;
- AA<sub>3</sub> is a neutral/nonpolar/large/cyclic or neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- AA<sub>4</sub> is a neutral/polar/small amino acid residue;
- 30 AA<sub>5</sub> is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;
- AA<sub>6</sub> is a neutral/nonpolar/large/non-cyclic/neutral/polar/large/non-cyclic amino acid residue;
- AA<sub>7</sub> is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- AA<sub>8</sub> is a neutral/polar/small amino acid residue
- AA<sub>9</sub> is a neutral/polar/small amino acid residue;
- 35 Z<sub>1</sub> is hydrogen, amino, acetyl or at least one amino acid residue or the desamino form thereof; and
- Z<sub>2</sub> is hydroxyl, carboxyl, non-amino acids such as agmatine, or at least one amino acid residue, including carboxamide or alkylamide forms thereof.

The most preferred sequence of this core nonapeptide is W-S-P-C-S-V-T-C-G (SEQ ID No. 1). Other preferred embodiments include polypeptide compounds wherein:

- 40 AA<sub>1</sub> is tryptophan or n-formyl-tryptophan;
- AA<sub>2</sub> is serine, threonine or aspartic acid;
- AA<sub>3</sub> is proline, glutamic acid, serine or isoleucine;
- AA<sub>4</sub> is cysteine;
- AA<sub>5</sub> is serine or asparagine;
- 45 AA<sub>6</sub> is valine or threonine;
- AA<sub>7</sub> is threonine or serine;
- AA<sub>8</sub> is cysteine;
- AA<sub>9</sub> is glycine or serine, including carboxamide forms thereof.

Particularly preferred are those embodiments wherein the sequence is selected from the group  
50 consisting of:

55

	<u>SEQ ID</u> <u>No.</u>	<u>Compound</u>	<u>Structure</u>
5	1	p1	W-S-P-C-S-V-T-C-G
		p5	fW-S-P-C-S-V-T-C-G-NH <sub>2</sub>
		p6	W-S-P-C-S-V-T-C-G-NH <sub>2</sub>
	2	p18	W-D-I-C-S-V-T-C-G
10	3	p19	W-S-S-C-S-V-T-C-G
	4	p20	W-T-S-C-S-T-S-C-G
	5	p11	W-S-P-W-S-E-W-T-S-C-S-T-S-C-G-N-G-I-Q-Q-R-G-R
	6	p17	W-S-H-W-S-P-W-S-S-C-S-V-T-C-G-D-G-V-I-T-R-I-R
15	7	p12	W-G-P-W-S-P-W-D-I-C-S-V-T-C-G-G-G-V-Q-K-R-S-R
	8		W-S-P-C-S-V-T-C-S
	9		W-S-Q-C-S-V-T-C-G
20	10		W-S-Q-C-N-V-T-C-G
	11		W-T-P-C-S-V-T-C-G
	13	p21	W-D-E-C-R-Q-T-C-G-A-S
	14	p22	W-S-A-C-S-L-G-C-D
25		p25	W-D-I-C-S-V-T-C-G-NH <sub>2</sub>
		p26	W-S-S-C-S-V-T-C-G-NH <sub>2</sub>
		p27	W-T-S-C-S-T-S-C-G-NH <sub>2</sub>
30	12	p4	D-G-G-W-S-H-W-S-P-W-S-S-C-S-V-T-C-G-D-G-V-I-T- R-I-R-L-C-N-S-P-S-P-Q-M-N-G-K-P-C-E-G-E-A-R- E-T-K-A-C-K-K-D-A-C-P-I-N-G-G

35 Compounds within the scope of the present invention can be synthesized chemically by means well known in the art such as, e.g., solid phase peptide synthesis. The synthesis is commenced from the carboxy-terminal end of the peptide using an alpha-amino protected amino acid. t-Butylocarbonyl (Boc) protective groups can be used for all amino groups even though other protective groups are suitable. See  
 40 Stewart et al., "Solid-Phase Peptide Synthesis," W. H. Freeman Co., San Francisco (1969) and Merrifield, J. Am. Chem. Soc. 85: 2149-2154 (1963). These and other methods of peptide synthesis are also exemplified by U.S. Patents Nos. 3,862,925, 3,842,067, 3,972,859 and 4,105,602.

Conveniently, compounds may be synthesized using manual techniques or automatically employing, for example, an Applied BioSystems 430A Peptide Synthesizer (Foster City, California) or a Biosearch SAM II  
 45 automatic peptide synthesizer (Biosearch, Inc., San Rafael, California), following the instructions provided in the instruction manual supplied by the manufacturer.

It will be readily appreciated by those having ordinary skill in the art of peptide synthesis that the intermediates which are constructed in accordance with the present disclosure during the course of synthesizing the present compounds are themselves useful compounds and are thus within the scope of the  
 50 invention.

Alternatively, selected compounds of the present invention can be produced by expression of recombinant DNA constructs prepared in accordance with well-known methods. Such production can be desirable to provide large quantities or alternative embodiments of such compounds.

#### 55 C. Administration

Compounds of the present invention have thrombospondin-like activity in the intact animal. Compounds of the present invention and compositions containing them which are shown to have the physiological effect

of inhibiting or mimicing the effect of intact thrombospondin find use in numerous therapeutic and prophylactic applications, such as cancer therapy, wound healing, atherosclerosis, thrombotic or thrombolytic conditions, angiogenesis, complement activation, or cell attachment.

Thus the present invention also provides compositions containing an effective amount of compounds of the present invention, including the nontoxic addition salts, amides and esters thereof, which may, alone, serve to provide the above-recited therapeutic benefits. Such compositions can also be provided together with physiologically tolerable liquid, gel or solid diluents, adjuvants and excipients.

These compounds and compositions can be administered to animals for veterinary use, such as with domestic animals, and clinical use in humans in a manner similar to other therapeutic agents. In general, the dosage required for therapeutic efficacy will range from about 1  $\mu$ g to 300 mg/kg, more usually 10  $\mu$ g to 30 mg/kg of the host body weight. Alternatively, dosages within these ranges can be administered by constant infusion over an extended period of time, usually exceeding 24 hours, until the desired therapeutic benefits have been obtained.

Typically, such compositions are prepared as injectables, either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be emulsified. The active ingredient is often mixed with diluents or excipients which are physiologically tolerable and compatible with the active ingredient. Suitable diluents and excipients are, for example, water, saline, dextrose, glycerol, or the like, and combinations thereof. In addition, if desired, the compositions may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, stabilizing or pH buffering agents, and the like.

The compositions are conventionally administered parenterally, by injection, for example, either subcutaneously or intravenously. Additional formulations which are suitable for other modes of administration include suppositories, intranasal aerosols, and, in some cases, oral formulations. For suppositories, traditional binders and excipients may include, for example, polyalkylene glycols or triglycerides: such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1%-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, cellulose, magnesium carbonate, and the like. These compositions take the form of solutions, suspensions, tablets, pills, capsules, sustained release formulations, or powders, and contain 10%-95% of active ingredient, preferably 25%-70%. These oral formulations include formulations designed to protect the peptide until it can be absorbed.

The peptide compounds may be formulated into the compositions as neutral or salt forms. Pharmaceutically acceptable non-toxic salts include the acid addition salts (formed with the free amino groups) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

In addition to the compounds of the present invention which display thrombospondin-like activity, compounds of the present invention can also be employed as intermediates in the synthesis of such useful compounds.

The compounds of the invention can be homopolymerized to themselves (i.e., (peptide)<sub>n</sub>) or, heteropolymerized to one another (i.e., (peptide 1-peptide 2)). The compounds can also be cyclized through disulfide or other means. The compounds can also be conjugated to biocompatible polymeric compounds, such as BIOPOLTM (W. R. Grace & Co.-Conn.)

While not wishing to be bound by any theory, it is believed that the compositions of the invention act as agonists or antagonists to native thrombospondin. These compounds are also believed to act as agonists or antagonists to circumsporozoite protein, thrombospondin related anonymous protein, and properdin complement protein. Further, since the compounds of the invention are small in size (relative to intact thrombospondin) the properties which they exhibit are more likely to be specific in nature, as opposed to the actions of other generally adhesive compounds such as RGD containing compounds (the sequence of which is found in over a hundred proteins) and fibronectin. The side effects of the peptide compounds of the invention are greatly reduced when compared with these broadly adhesive compounds.

#### 55 D. Use

As stated previously, the compounds of the invention can be used in a variety of biological, prophylactic or therapeutic areas. It is contemplated that these compounds are useful in prevention or treatment of any



disease state or conditions wherein thrombospondin-like activity plays a role. These disease states and conditions include, but are not limited to, metastasis, wound healing, atherosclerosis, thrombotic conditions, thrombolytic conditions, angiogenesis, cell proliferation, and complement activation. Antibodies directed against the compounds of the invention are also useful as diagnostic reagents, therapeutics, or carriers of other compounds.

Numerous in vitro and in vivo assays can be used to demonstrate compounds having thrombospondin-like activity. These assays include, but are not limited to, cell adhesion assays, platelet aggregation assays and cell proliferation assays.

## 10 METASTASIS

Metastasis is the spread of disease from one part of the body to another unrelated to it, as in the transfer of the cells of a malignant tumor by way of the bloodstream or lymphatics. It is believed that metastasis is effected through a cascade mechanism which includes adhesion of tumor cells to endothelium, retraction of the endothelium, matrix degradation of the basement membrane and invasion of the tumor cells into the bloodstream. Intervention at any phase in this cascade could be beneficial to the treatment or prevention of metastatic cancers.

As mentioned earlier, native thrombospondin has been shown to potentiate tumor cell metastasis. The mechanisms by which the thrombospondin potentiation occurs are not presently well understood.

Antimetastasis activity is characterized by the ability of the compounds to bind to melanoma cells in vitro (Tuszynski et al., Anal. Bio. (1990) 184:189-91), and the ability to reduce the size and number of tumor colonies in vivo (Tuszynski et al. Cancer Research (1987) 47:4130-4133).

The compounds of this invention are useful as antimetastatic agents, particularly useful as anti-pulmonary metastatic agents. These compounds inhibit the adhesion of metastatic tumor cells, particularly those which are responsive to thrombospondin. The compounds also reduce tumor colony number as well as tumor colony size.

There are a number of mechanisms by which such antimetastatic activity can be occurring. The peptides can be cytotoxic, or inhibit cell proliferation. As inhibitors of cell proliferation, the compounds can act to 1) inhibit mitogenesis, 2) inhibit angiogenesis, or 3) activate the complement pathway and the associated killer cells.

The compounds of the invention can also find use in biomedical devices. Since the compounds have the ability to promote the attachment of metastatic tumor cells, it is possible to coat a biomedical device with the compounds to effect the removal of circulating tumor cells from blood or lymph. The biomedical device is also useful to trap hepatomas.

Another use of the compounds is as a carrier to target toxins, drugs, hormones or imaging agents to metastatic tumor cells for diagnostic or therapeutic purposes. These carriers would also bind to hepatomas.

## WOUND HEALING

Wound healing is the closure of wounds and can be divided into four essential components: inflammation, angiogenesis, collagen deposition and epithelialization. All four components play a role in the healing of wounds.

Wound healing activity is characterized by the ability of the compounds to show angiogenic activity, the ability of the compounds to stimulate collagen deposition and DNA synthesis in the in vivo sponge model or the ability of the compounds to improve wound healing or reduce healing time in an in vivo partial or full thickness wound model.

## ATHEROSCLEROSIS

Atherosclerosis is a disease state which is characterized by the deposition of small fatty nodules on the inner walls of the arteries, often accompanied by degeneration of the affected areas.

Atherosclerosis activity is characterized by the capacity of the compounds to inhibit the development of aortic lesions in rabbits fed a high cholesterol diet. Other assays which characterize atherosclerosis activity include regulating vascular smooth muscle and endothelial cell proliferation, contractile phenotype and response to injury. These assay procedures are well established in the art.

## THROMBOTIC CONDITIONS

5 The thrombotic activity associated with the compounds of the invention acts to inhibit platelet aggregation and platelet thrombus formation. Platelets participate in blood coagulation via binding fibrinogen, platelet aggregation and thrombus formation. As anti-thrombotics, these peptides can be useful in the following conditions: myocardial infarction, thromboembolic disease and thrombotic complications due to cancer and cardiovascular disease.

10 The compounds of this invention have the ability to specifically inhibit the second stage of platelet aggregation (i.e., the thrombospondin dependent stage of platelet aggregation). This activity allows the compounds to be useful in inhibiting thrombocytopenia caused as a result of disease state (i.e., Gray Platelet Syndrome, Essential Thrombocythemia, Myeloproliferative Syndrome) or induced by therapy (i.e., cancer therapy) or autoimmune diseases. These compounds also act to prevent coronary artery reocclusion following balloon catheterization.

15 The compounds of this invention modulate the formation and structure of blood clots. Thrombospondin is incorporated into fibrin clots and serves as a substrate for blood clotting Factor XIIIa. An IQQ sequence motif in thrombospondin has been implicated in crosslinking to factor XIIIa. Peptides containing IQQ modulate structure and formation of clots. Known fibrinolytic *in vitro* assays demonstrate this ability.

20 Antithrombotic activity is characterized by a number of assays, including 1) inhibition of ADP or thrombin-induced platelet aggregation in washed platelets; 2) inhibition of platelet aggregation in platelet-rich plasma; 3) inhibition of collagen induced platelet aggregation measured *in vivo*; and 4) inhibition of induced thrombus formation in a carotid artery - in this assay the peptide would delay or prevent occlusion of the artery following thrombus induction.

Alternatively, the compounds of the present invention are useful as potent clotting agents. The effect can be localized and is long lasting. This activity is useful when clotting is necessary (i.e., surgery or in hemophilia).

## 25 THROMBOLYTIC CONDITIONS

The thrombolytic activity associated with the compounds of the invention act to alter the structure of a formed thrombus, i.e., dissolution of a blood clot. Thrombolytic activity is characterized as the ability to enhance the dissolution of fibrin in the presence of plasmin (i.e., standard clot lysis assay).

## 30 ANGIOGENESIS

35 Angiogenesis is the formation of blood and lymph vessels. The compounds of this invention are useful in the modulation of angiogenesis, particularly in enhancing wound healing, inhibiting or preventing tumor growth and metastasis, diabetic retinopathy, neovascular glaucoma, and rheumatoid arthritis. Standard angiogenesis assays are well known in the art. These assays include, but are not limited to, proliferation and migration studies using various cell lines, collagenase inhibition and *in vivo* neovascularization on chicken chorioallantoic membranes (CAM assay).

## 40 COMPLEMENT ACTIVITY

45 The complement activity associated with the compounds of the invention can play a role in a variety of disease states. The peptides enhance complement-mediated clearance and inactivation mechanisms in both natural and acquired resistance to infection. The complement activity of the peptides serves to promote tumoricidal activity of the complement protein C3b. Additionally, complement proteins are known to contribute to reperfusion injury following heart attacks, and the compounds of the invention can inhibit such activity and are thus useful to lessen heart tissue death and tissue injury during a heart attack.

## 50 ANTIVIRAL

Viral infections involve attachment to cells of viral particles. The peptides of this invention interfere with viral attachment and are thus useful as antiviral agents. The peptides of this invention are also useful as anti-bacterials.

## 55 ANTIBODIES

Compounds of the present invention can also be used for preparing antisera for use in immunoassays employing labelled reagents, usually antibodies. Conveniently, the polypeptides can be conjugated to an

antigen by means of dialdehydes, particularly from 4 to 6 carbon atoms and aliphatic, or carbodimide. These compounds and immunologic reagents may be labelled with a variety of labels such as chromophores, fluorophores such as, e.g., fluorescein or rhodamine, radioisotopes such as  $^{125}\text{I}$ ,  $^{35}\text{S}$ ,  $^{14}\text{C}$ , or  $^3\text{H}$ , or magnetized particles, by means well known in the art.

5 These labelled compounds and reagents, or labelled reagents capable of recognizing and specifically binding to them, can find use as, e.g., diagnostic reagents. Samples derived from biological specimens can be assayed for the presence or amount of substances having a common antigenic determinant with compounds of the present invention.

Thrombospondin levels are elevated in the serum of patients with metastatic breast and colon cancer (Tuszynski et al., Antithrombotic Therapy (1989) A28 and Smith et al., American Association of Clinical Oncology (in press)). Antibodies against the peptides of the invention can be useful as reagents in diagnostic/prognostic assays for various types of cancer, including but not limited to, gastrointestinal tract (gastric, colonic, and rectal) carcinomas, breast carcinomas and hepatic carcinomas.

In addition, monoclonal antibodies can be prepared by methods known in the art. The polyclonal and 15 monoclonal antibodies can find therapeutic use in a number of cancer therapies. First, the antibodies can be used to sequester thrombospondin. This is useful since thrombospondin mediates tumor cell metastasis. Second, the antibodies can be used to block thrombospondin present on the tumor cell surface. Third, cytotoxic drugs, hormones, or imaging agents can be coupled to the antibodies for use in cancer therapy. Fourth, a biomedical device can be coated with the antibodies to remove excess thrombospondin from 20 serum or to remove cells which bear thrombospondin on the cell surface.

The peptides of the invention can also be used to isolate thrombospondin cell surface receptors from extracts of cells or cell membranes. The thrombospondin cell surface receptors can be used to develop better thrombospondin analogs or to remove excess thrombospondin from serum.

The following examples are provided by way of illustration, rather than implying any limitation of the 25 subject matter.

#### EXAMPLES

The peptides of this invention can be synthesized by conventional methods of peptide synthesis. A 30 preferred method is the solid phase synthesis of Merrifield, J. Amer. Chem. Soc. 85, 2149-2154 (1963); Science 150, 178-185 (1965); Ibid. 232, 341-347 (1986). Solid phase synthesis is generally initiated from the C-terminal of the peptide by coupling a protected alpha amino acid to a suitable resin, e.g., phenylacetamidomethyl (PAM) polystyrene resin, or p-methylbenzhydrylamine (MBHA) resin when synthesizing a peptide with a C-terminal carboxamide. In the present invention, the peptides were synthesized 35 by solid-phase techniques performed on an Applied Biosystems 430A Peptide Synthesizer (Foster City, California) using t-butyloxycarbonyl (t-Boc) alpha amino-group protected amino acids in accordance with the instructions of the manufacturer. During this synthesis, suitable amino acid side-chain protecting groups are used as needed. Thus, aspartic acid is protected on the beta-carboxyl group as the benzyl ester and arginine is protected on the guanidino group by tosyl. After the desired peptide has been synthesized, the 40 peptide is cleaved from the resin and protecting groups are removed by treatment with a reagent such as hydrogen fluoride (HF). The peptide can then be purified by high performance liquid chromatography (HPLC) or other such methods of peptide purification. Background information on the established procedures for solid phase peptide synthesis can be found in "Solid Phase Peptide Synthesis" by Stewart and Young, W. H. Freeman & Co., San Francisco, 1969.

45 In accordance with the above description, the following procedures were used for the chemical synthesis of novel synthetic peptides:

##### Procedure A

50 0.1 mmole of selected Boc-AA<sub>n</sub>-OCH<sub>2</sub>-PAM Resin (0.2-0.8 mmole/g resin) (Applied Biosystems, Inc.) or p-mBHA Resin (0.2-0.7 mmole/g resin) (Applied Biosystems, Inc.) is treated according to Schedule A for the incorporation of the Boc-AA<sub>n-1</sub> or Boc-AA<sub>n</sub>, respectively.

##### Schedule A. Small-scale Rapid Cycle Chemistry

- 55
1. 5-minute neat TFA wash
  2. 40s DMF flow wash
  3. 1-minute treatment with 20% DIEA in DMF
  4. 40s DMF flow wash

alents of preformed symmetric anhydride of a suitable protected t-boc amino  
 iod

Boc-AA<sub>n</sub>-OCH<sub>2</sub>-PAM Resin (0.2-0.8 mmole/g resin) (Applied Biosystems, Inc.) or  
 ole/g resin) (Applied Biosystems.) is treated according to Schedule B for the  
 1 or Boc-AA<sub>n</sub> respectively.

IP/HOBT Chemistry  
 % TFA in DCM.  
 3% TFA in DCM.

: DIEA in DCM.  
 : DIEA in NMP.

ents of HOBT-ester of a suitably protected t-boc amino acid dissolved in NMP.  
 iod.

0% and subsequent 16-minute coupling period.  
 1. nts DIEA and subsequent 7-minute coupling period.

: anhydride, 5% DIEA in DCM for 2 minutes.  
 : anhydride in DCM for 4 minutes.

#### ID No. 1) WSPCSVTCG

t-boc-Gly-OCH<sub>2</sub> PAM resin (0.79 mmol/g) was subjected to the following  
 ly protected amino acids: t-boc-Cys(4-CH<sub>3</sub>Bzl), t-boc-Thr(Bzl), t-boc-Val, t-boc-  
 l), t-boc-Pro, t-boc-Ser(Bzl), and t-boc-Trp. Resultant dry, N-terminal protected  
 -d in anhydrous hydrogen fluoride containing 5% anisole, 5% dimethyl sulfide,  
 hours at -5 °C. HF and volatile scavengers were removed with nitrogen sparge  
 e was suspended in cold diethyl ether. The peptide-resin mixture was washed  
 er, then the peptide was extracted with 30% acetic acid. This solution was  
 ophilized to afford the crude peptide. Purification was achieved on an Amicon  
 izing reverse phase chromatography employing 0.1% aqueous trifluoroacetic  
 on afforded by the addition of 0.1% TFA in 90% acetonitrile and 10% H<sub>2</sub>O.  
 d pooled based on a purity of 90%, as judged by analytical reverse-phase  
 e diluted with deoxygenated H<sub>2</sub>O and lyophilized to afford the pure peptide as a

#### CSVTCG-NH<sub>2</sub>

ethylbenzylhydramine resin (0.62 mmol/g) was subjected to the following  
 ly protected amino acids: t-boc-Gly, t-boc-Cys(4-CH<sub>3</sub>Bzl), t-boc-Thr(Bzl), t-boc-  
 s(4-CH<sub>3</sub>Bzl), t-boc-Pro, t-boc-Ser(Bzl), t-boc-Trp(CHO). The resultant n-terminal  
 tidyl-resin was dried then suspended in anhydrous HF containing 8% anisole  
 eatment was for 0.5 hours at -20 °C and 2 hours at 0 °C. The HF was removed  
 eptide-resin mixture was suspended in and washed 3 times with diethyl ether.  
 with 25% acetic acid. The resin was washed with 50% acetic acid and with H<sub>2</sub>O.

5. Addition of 1-10 equivalents of preformed symmetric anhydride of a suitable protected t-boc amino acid dissolved in DMF
6. 10-minute coupling period
7. 40s DMF flow wash

5 Procedure B

0.5 mmole of selected Boc-AA<sub>n</sub>-OCH<sub>2</sub>-PAM Resin (0.2-0.8 mmole/g resin) (Applied Biosystems, Inc.) or p-mBHA Resin (0.2-0.7 mmole/g resin) (Applied Biosystems, Inc.) is treated according to Schedule B for the incorporation of the Boc-AA<sub>n-1</sub> or Boc-AA<sub>n</sub> respectively.

10

Schedule B. Large-scale NMP/HOBT Chemistry

1. 3-minute wash with 30% TFA in DCM.
2. 17-minute wash with 50% TFA in DCM.
3. Wash 5X with DCM.
- 16 4. 1-minute wash with 5% DIEA in DCM.
5. 1-minute wash with 5% DIEA in NMP.
6. Wash 5X with NMP.
7. Addition of 1-4 equivalents of HOBT-ester of a suitably protected t-boc amino acid dissolved in NMP.
8. 30-minute coupling period.
- 20 9. Addition of DMSO to 20% and subsequent 16-minute coupling period.
10. Addition of 3.8 equivalents DIEA and subsequent 7-minute coupling period.
11. Wash 3X with DCM.
12. Wash with 10% acetic anhydride, 5% DIEA in DCM for 2 minutes.
13. Wash with 10% acetic anhydride in DCM for 4 minutes.
- 25 14. Wash 4X with DCM.

Example 1

Chemical Synthesis of (SEQ ID No. 1) WSPCSVTGG

30 Compound p1

Briefly, 0.13 grams of t-boc-Gly-OCH<sub>2</sub> PAM resin (0.79 mmol/g) was subjected to the following sequential addition of suitably protected amino acids: t-boc-Cys(4-CH<sub>3</sub>Bzl), t-boc-Thr(Bzl), t-boc-Val, t-boc-Ser(Bzl), t-boc-Cys(4-CH<sub>3</sub>Bzl), t-boc-Pro, t-boc-Ser(Bzl), and t-boc-Trp. Resultant, dry, N-terminal protected  
 35 peptidyl-resin was suspended in anhydrous hydrogen fluoride containing 5% anisole, 5% dimethyl sulfide, and 1% p-thiocresol for two hours at -5 °C. HF and volatile scavengers were removed with nitrogen sparge and the peptide-resin mixture was suspended in cold diethyl ether. The peptide-resin mixture was washed three times with diethyl ether, then the peptide was extracted with 30% acetic acid. This solution was diluted 1:1 with H<sub>2</sub>O and lyophilized to afford the crude peptide. Purification was achieved on an Amicon  
 40 C18 MC-250-10 column utilizing reverse phase chromatography employing 0.1% aqueous trifluoroacetic acid with gradient generation afforded by the addition of 0.1% TFA in 90% acetonitrile and 10% H<sub>2</sub>O. Fractions were collected and pooled based on a purity of 80%, as judged by analytical reverse-phase HPLC. Pooled fractions were diluted with deoxygenated H<sub>2</sub>O and lyophilized to afford the pure peptide as a trifluoroacetic acid salt.

45

Example 2

Chemical Synthesis of WSPCSVTGG-NH<sub>2</sub>

50 Compound p6

Briefly, 0.17 g of p-methylbenzylhydramine resin (0.62 mmol/g) was subjected to the following sequential addition of suitably protected amino acids: t-boc-Gly, t-boc-Cys(4-CH<sub>3</sub>Bzl), t-boc-Thr(Bzl), t-boc-Val, t-boc-Ser(Bzl), t-boc-Cys(4-CH<sub>3</sub>Bzl), t-boc-Pro, t-boc-Ser(Bzl), t-boc-Trp(CHO). The resultant n-terminal  
 55 deprotected, formylated peptidyl-resin was dried then suspended in anhydrous HF containing 8% anisole and 2% dimethyl sulfide. Treatment was for 0.5 hours at -20 °C and 2 hours at 0 °C. The HF was removed with nitrogen sparge. The peptide-resin mixture was suspended in and washed 3 times with diethyl ether. The peptide was extracted with 25% acetic acid. The resin was washed with 50% acetic acid and with H<sub>2</sub>O.

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The aqueous solutions were pooled, diluted 1:1 with deoxygenated H<sub>2</sub>O and lyophilized to afford the formylated crude peptide. Deformylation was effected using the procedure described in Applied Biosystems 430A User Bulletin 18. (April 28, 1987). The peptide (5mg/ml concentrate) in 6M guanidine-HCl was cooled to 0 °C and ethanolamine added to 1 M. The pH is lowered to 6 with concentrated hydrochloric acid.

5 Purification was on an Amicon C<sub>18</sub> MC-250-10 column using reverse-phase chromatography. The ratio of acetonitrile to H<sub>2</sub>O was increased maintaining 0.1% TFA to achieve gradient elution. Collected fractions with a purity of 90%, as determined by analytical reverse-phase PHLC, were pooled, diluted with deoxygenated H<sub>2</sub>O and lyophilized to afford pure peptide as a trifluoroacetic acid salt.

### 10 Example 3

#### Chemical Synthesis of fWSPCSVTCG-NH<sub>2</sub>

#### Compound p5

15

Briefly, procedure was same as for Example 2 except the deformylation step is not performed. The afforded crude peptide obtained past HF cleavage and subsequent lyophilization was purified as the formylated, C-terminal amide species.

### 20 Example 4

#### Other Thrombospondin Fragments or Analogs

25 Following the procedures outlined in Examples 1, 2 and 3 with appropriate modification, the following thrombospondin fragments or analogs were synthesized:

	SEQ ID		
	No.	Compound	Structure
30	2	p18	W-D-I-C-S-V-T-C-G
	3	p19	W-S-S-C-S-V-T-C-G
	4	p20	W-T-S-C-S-T-S-C-G
35	5	p11	W-S-P-W-S-E-W-T-S-C-S-T-S-C-G-N-G-I-Q-Q-R-G-R
	6	p17	W-S-H-W-S-P-W-S-S-C-S-V-T-C-G-D-G-V-I-T-R-I-R
	7	p12	W-G-P-W-S-P-W-D-I-C-S-V-T-C-G-G-G-V-Q-K-R-S-R
	8		W-S-P-C-S-V-T-C-S
40	9		W-S-Q-C-S-V-T-C-G
	10		W-S-Q-C-N-V-T-C-G
	11		W-T-P-C-S-V-T-C-G
45	13	p21	W-D-E-C-R-Q-T-C-G-A-S
	14	p22	W-S-A-C-S-L-G-C-D
		p25	W-D-I-C-S-V-T-C-G-NH <sub>2</sub>
		p26	W-S-S-C-S-V-T-C-G-NH <sub>2</sub>
50		p27	W-T-S-C-S-T-S-C-G-NH <sub>2</sub>
	12	p4	D-G-G-W-S-H-W-S-P-W-S-S-C-S-V-T-C-G-D-G-V-I-T-R-
			I-R-L-C-N-S-P-S-P-Q-M-N-G-K-P-C-E-G-E-A-R-E-T-K-
55			A-C-K-K-D-A-C-P-I-N-G-G

Exempl 5Direct Adhesion Assay

5 It is believed that thrombospondin acts in metastasis through its adhesive properties. An assay was developed, generally in accordance with the disclosure of Tuszynski et al. (Anal. Bio. (1990) 184:189-91) which evaluates the ability of melanoma cells to adhere to the thrombospondin fragments or analogs of the invention. In this assay, wells of a 96-well microtiter dish (Costar, Cambridge, Massachusetts) were incubated for two to three hours with 50  $\mu$ l of a 40  $\mu$ g/ml solution of various ligands in 20 mM NaCl, pH 7.3.

10 Thrombospondin (purified by the method of Tuszynski et al., J. Biol. Chem. (1985) 260:12240-5), fibronectin (Sigma Chemical Co., Missouri) served as the positive control. Bovine serum albumin (BSA) (Sigma Chemical Co.) served as the negative control. Thrombospondin analogs of the invention (SEQ ID No. 1) p1, p5 and p6 were synthesized as described in Examples 1 through 3. Following peptide adhesion to the microtiter dish, the wells were aspirated, treated with 200  $\mu$ l phosphate buffered saline (PBS) containing 1% BSA for 1 hour and then washed three more times with 200  $\mu$ l PBS.

Mouse B16F10 melanoma cells were grown and harvested during log phase of growth using standard procedures. The harvested cells were washed two times in serum-free Dulbecco's minimum essential medium (DMEM) (Flow Laboratories) and suspended in DMEM at a final concentration of  $4 \times 10^5$  cells/ml. Of the cell suspension 100  $\mu$ l was added to each well of the microtiter dish containing the various ligands and the dish incubated at 37°C in a CO<sub>2</sub> incubator for 1 hour. Nonadherent cells were removed by aspiration and the wells washed three times with 200  $\mu$ l of PBS. The total cell-associated protein was determined by dissolving the attached cells directly in the microtiter wells with 200  $\mu$ l of the Pierce BCA working solution (Pierce Chem. Co. Booklet No. 23225 (1987)). The plate was covered with an adhesive mylar sheet (Flow Labs) and incubated at 60°C for 30 minutes. Plates were allowed to cool to room

25 temperature, cover sheets were removed, and the absorbance of each well was determined at 562 nm with a microtiter plate reader (Biotek, Burlington, Vermont.)

The results shown in Table 1 indicate that the peptides of the inventions p1, p5, and p6 display adhesive properties.

Table 1

<u>Absorbed Compound (2 <math>\mu</math>g)</u>	<u>Adhesion as % Thrombospondin</u>
Thrombospondin	100
p1 (SEQ ID No. 1)	98
p5	79
p6	70
BSA	13

Example 6Competitive Assay to Measure Thrombospondin Specific Adhesion to Melanoma Cells

50 The direct adhesion assay of Example 5 was modified to measure the ability of the peptide compounds of the invention to compete with intact thrombospondin for adhesion to melanoma cells.

In this assay, intact thrombospondin is absorbed onto microtiter dishes as described in Example 5. The assay is similar to Example 5 except that before the melanoma cells are added to the microtiter dishes they are preincubated for 15 minutes with 100  $\mu$ g/ml of various ligands.

55 The results shown in Table 2 indicate that peptide p1 (SEQ ID No. 1) is able to effectively compete with thrombospondin for adhesion of melanoma cells. Peptides p5 and p6 compete to a lesser degree at this concentration.



Table 2

	<u>Compound</u>	<u>Concentration</u> <u>(<math>\mu</math>g/ml)</u>	<u>% of buffer control</u>
5	buffer	---	100
	p1 (SEQ ID No. 1)	100	20
		100	42
10		50	40
		10	49
15	p5	100	62
		101	108
		50	104
20		10	99
		100	83
		200	89
25		300	83
30	p6	100	89
		200	74
		300	74

35 Example 7Direct Platelet Adhesion Assay

40 The number of adherent platelets was essentially determined as previously described (Tuszynski et al., Anal. Bio. supra). Briefly, 100  $\mu$ l of  $5 \times 10^8$  platelets/ml washed as previously described (Tuszynski, et al., Blood 72, 109-225, 1988) were added to microtiter plates, the wells of which were treated with 50  $\mu$ l of a 40  $\mu$ g/ml peptide or protein solution (Hepes buffered saline, pH 7.4). Solutions were dried at room temperature after incubation in a fume hood for two hours. Wells were blocked for one hour with 1% BSA. Platelets (100  $\mu$ l) were incubated in the wells for 30 minutes and non-adherent platelets were removed by aspiration. Wells 45 were washed 3X with 200  $\mu$ l of Hepes buffered saline, pH 7.4. The number of adherent platelets was determined by measuring the platelet-derived protein using the BCA protein assay.

The results shown in Table 3 indicate that at this concentration p1 (SEQ ID No. 1), p6, and to a lesser degree, p5 adhere to platelets.

50

55

Table 3

	<u>Compound</u>	<u>Adherence as % Control</u>
5	Thrombospondin	100
	BSA	0.7
	p1 (SEQ ID No. 1)	39
10	p5	2.1
	p6	38
	fibronectin	83

15

Example 8Competitive Assay to Measure Thrombospondin Specific Adhesion to Platelets

20

The direct adhesion assay of Example 7 was modified to measure the ability of the peptide compounds of the invention to compete with intact thrombospondin or fibronectin for adhesion of platelets.

In this assay, thrombospondin (TSP) or fibronectin (FN) was absorbed onto microtiter dish as described in Example 7. The assay was similar to Example 7 except that before platelets were added to the microtiter dishes they were preincubated with 250  $\mu$ g/ml of various ligands.

The results shown in Table 4 indicate that peptide p1 (SEQ ID No. 1) effectively competed with thrombospondin for platelet adhesion, but did not compete with fibronectin.

30

Table 4

	<u>Compound</u>	<u>Concentration (<math>\mu</math>g/ml)</u>	<u>Absorbed Protein</u>	<u>% Buffer Control</u>
35	buffer	---	TSP	100
	p1 (SEQ ID No. 1)	250	TSP	71
	p5	250	TSP	98
40	buffer	---	FN	100
	p1 (SEQ ID No. 1)	250	FN	108
	p5	250	FN	109
45	p6	250	FN	96

45

Example 9Platelet Aggregation Assay

50

Platelet aggregation was monitored on a single-channel aggregometer equipped to measure luminescence (Chromo-Log, Havertown, Pennsylvania). Platelet-rich-plasma (PRP) was obtained from whole blood anti-coagulated with 0.38% Sodium Citrate by centrifugation at 150 X g for 20 minutes. Peptides were added to 0.5 ml of PRP and aggregation initiated with 1  $\mu$ M ADP. Buffer blanks contained diluent (water) but no peptide.

The results shown in Figure 1 indicate that p1 (SEQ ID No. 1), p5 and p6 inhibit platelet aggregation.

Example 10Antimetastatic Activity In Vivo

- 5 The in vivo model used to demonstrate the antimetastatic activity of the peptide compounds of the invention is described by Tuszyński et al. (*Cancer Res.* (1987) 47:4130-4133). Briefly, C57 black mice were intravenously injected with  $5 \times 10^5$  B<sub>16</sub>F<sub>10</sub> mouse melanoma cells in the presence of either control buffer (Hepes buffered saline, pH 7.4), or the indicated amount of peptide compound of the invention p1 (SEQ ID No. 1), p5 or p6. After 16 days, the mice were sacrificed and the number of lung tumors counted.
- 10 The results shown in Table 5 indicate the peptides of the invention have antimetastatic activity. The p1-, p5- and p6-treated animals developed statistically fewer tumors than controls. Additionally, the lung tumors were smaller in size in the p1- and p6-treated animals.

Table 5

	<u>Compound</u>	<u>Concentration</u> <u>µg/mouse</u>	<u>% buffer control</u>
	buffer	---	100
20	p1 (SEQ ID No. 1)	30	49
	p5	30	87
	p6	30	91
25	p6	100	102
	p6	300	50

30

Example 11CAM Assay

- 35 Assays for antiangiogenic properties on the chicken chorioallantoic membrane (CAM) were performed using the "egg culture" technique (Auerbach, *Dev. Biol.* (1974) 41:391 and Crum, *Science* (1985) 230:1375). The fertilized White Leghorn egg (Truslow Farms, Chestertown, MD) used in this method was removed from the shell on day 3 (after fertilization) and placed in a culture dish. The "cultured egg" was maintained through day 6 at 37° C in an atmosphere of 2% CO<sub>2</sub> and a relative humidity of 70%. After day 6
- 40 incubation was carried out without supplemental CO<sub>2</sub>.

- Test materials were implanted on the CAM using low gelling temperature agarose pellets (Castellot, *J. Cell Physiol.* (1986) 127:323). This assay was performed twice. In the first assay, the test materials included pellets containing 90, 9, and 0.9 µg of peptide p1 (SEQ ID No. 1) diluted in water. In the second assay, the test materials included pellets containing 100 µg peptide p1 diluted in water, 20, 50, 100, and 200 µg of
- 45 peptide p1 diluted in phosphate buffered saline. A positive control which was a pellet containing 60 µg hydrocortisone and 50 µg heparin was used in both assays. The sample pellets were placed on day 6 CAMs. Eighteen to twenty-one pellets were implanted for each test sample. Only one sample per egg was employed due to the possible cumulative effects of sample toxicity to the embryo.

- Antiangiogenesis was examined in an assay for inhibition of embryonic angiogenesis as described by
- 50 Fenselau in "Growth and Maturation Factors" (1984) 2:175. After 1 or 2 days the samples were evaluated for their antiangiogenic effects on the CAM. Antiangiogenesis scores range from 0 (for no change in vessel growth around the implant), to 1 (for a reduction in capillary number around the implant), and to 2 (for the absence of all capillaries around the implant). These scores permit the calculation of an Avascular Index [AVI]:

55

$$AVI = \frac{ax0 + bx1 + cx2}{a + b + c} \quad [MAX. \text{ VALUE} = 2.0]$$

where

a = Number of CAM with 0 Rating

b = Number of CAM with 1 Rating

c = Number of CAM with 2 Rating

a + b + c = Total Alive - Total Pellet Free

[Total Number of Usable Embryos at End of Test]

The value of the AVI is used as a primary indicator of antiangiogenic activity. The correlation between the range of AVI values and activity are indicated below:

<u>AVI Range</u>	<u>Level of Antiangiogenic Activity</u>
Below 0.5	Inactive
0.5 - 0.75	Weakly Active
0.75 - 1.0	Moderately Active
1.0 - 1.5	Highly Active
Above 1.5	Exceptionally Active

Inhibition of embryonic angiogenesis was also quantitated by measuring the size of the affected zone surrounding the pellet as well as the portion of the pellet that borders the affected region. The identity of the pellet contents was masked during the period of evaluation.

The results shown in Table 6 indicate that peptide p1 is weakly antiangiogenic. The study also showed no abnormal development or excessive embryo death at any dosage which indicates that p1 is not toxic. Additionally, on most of the CAMs, there was either reduced vascularity or avascularity around the pellet.

Table 6

	<u>Sample</u>	<u>AVI</u>	
		<u>Day 1</u>	<u>Day 2</u>
<u>Assay 1</u>	0.9 µg p1 in H <sub>2</sub> O	N.D.	0.46
	9 µg p1 in H <sub>2</sub> O	N.D.	0.46
	90 µg p1 in H <sub>2</sub> O	N.D.	0.77
	positive control	N.D.	1.06
<u>Assay 2</u>	20 µg p1 in PBS	N.D.	0.33
	50 µg p1 in PBS	N.D.	0.44
	100 µg p1 in PBS	0.80	0.40
	200 µg p1 in PBS	0.89	0.40
	100 µg p1 in H <sub>2</sub> O	0.83	0.59
	positive control	N.D.	1.33

Example 12Collagenase Inhibition Assay

5 Collagenase has been shown to have a key role in the angiogenic process (Langer et al., *Proc. Natl. Acad. Sci.* (1980) 77:4331; Thurgisser et al., *J. Natl. Cancer Inst.* (1982) 69:1049 and Rifkin et al. in *Pathology of the Endothelial Cell* (Academic Press, 1982) 191-197). Collagenase inhibitory activity was determined as described by Johnson-Witt (*Anal. Biochem.* (1980) 104:175). Peptides p1 (SEQ ID No. 1) and p27 were tested in the Johnson-Witt radiometric enzyme assay for collagenase activity at doses of 50  
10 ng, 100 ng, 250 ng, 500 ng, 1  $\mu$ g and 2  $\mu$ g. Peptide p27 was inhibitory (-21%) at the 2  $\mu$ g dose.

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SEQUENCE LISTING

INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Trp Ser Pro Cys Ser Val Thr Cys Gly  
 1 5

INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Trp Asp Ile Cys Ser Val Thr Cys Gly  
 1 5

INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Trp Ser Ser Cys Ser Val Thr Cys Gly  
 1 5

## INFORMATION FOR SEQ ID NO:4:

- 5 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

15 Trp Thr Ser Cys Ser Thr Ser Cys Gly  
 1 5

## 20 INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 23 amino acids  
 (B) TYPE: amino acid  
 25 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

## 30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Trp Ser Pro Trp Ser Glu Trp Thr Ser Cys Ser Thr Ser Cys Gly Asn  
 1 5 10 15  
 35 Gly Ile Gln Gln Arg Gly Arg  
 20

## 40 INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 23 amino acids  
 (B) TYPE: amino acid  
 45 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

## 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Trp Ser His Trp Ser Pro Trp Ser Ser Cys Ser Val Thr Cys Gly Asp  
 1 5 10 15  
 55 Gly Val Ile Thr Arg Ile Arg  
 20

INFORMATION FOR SEQ ID NO:7:

- 5 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 23 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear
- 10 (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
- 15 Trp Gly Pro Trp Ser Pro Trp Asp Ile Cys Ser Val Thr Cys Gly Gly  
 1 5 10 15  
 Gly Val Gln Lys Arg Ser Arg  
 20

INFORMATION FOR SEQ ID NO:8:

- 25 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear
- 30 (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
- 35 Trp Ser Pro Cys Ser Val Thr Cys Ser  
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INFORMATION FOR SEQ ID NO:9:

- 40 (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear
- 45 (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
- 50 Trp Ser Gln Cys Ser Val Thr Cys Gly  
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INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Trp Ser Gln Cys Asn Val Thr Cys Gly  
 1 5

INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:  
 (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Trp Thr Pro Cys Ser Val Thr Cys Gly  
 1 5

## INFORMATION FOR SEQ ID NO:12:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 60 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Asp Gly Gly Trp Ser His Trp Ser Pro Trp Ser Ser Cys Ser Val Thr  
 1                      5                      10                      15  
 Cys Gly Asp Gly Val Ile Thr Arg Ile Arg Leu Cys Asn Ser Pro Ser  
                     20                      25                      30  
 Pro Gln Met Asn Gly Lys Pro Cys Glu Gly Glu Ala Arg Glu Thr Lys  
                     35                      40                      45  
 Ala Cys Lys Lys Asp Ala Cys Pro Ile Asn Gly Gly  
                     50                      55                      60

## INFORMATION FOR SEQ ID NO:13:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 11 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Trp Asp Glu Cys Arg Gln Thr Cys Gly Ala Ser  
 1                      5                      10

## INFORMATION FOR SEQ ID NO:14:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 9 amino acids  
 (B) TYPE: amino acid  
 (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: peptide

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Trp Ser Ala Cys Ser Leu Gly Cys Asp  
 1                      5

## Claims

1. A method for promoting or inhibiting thrombospondin-like activity comprising administering an effective amount of a polypeptide compound comprising the formula:

$Z_1-AA_1-AA_2-AA_3-AA_4-AA_5-AA_6-AA_7-AA_8-AA_9-Z_2$

wherein:

- $Z_1$  is hydrogen, amino, acetyl, or at least one amino acid residue or the desamino forms thereof;
- $AA_1$  is a neutral/nonpolar/large/cyclic amino acid residue;
- $AA_2$  is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;
- $AA_3$  is a neutral/nonpolar/large/cyclic or neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- $AA_4$  is a neutral/polar/small amino acid residue;
- $AA_5$  is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;
- $AA_6$  is a neutral/nonpolar/large/non-cyclic/neutral/polar/large/non-cyclic amino acid residue;
- $AA_7$  is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- $AA_8$  is a neutral/polar/small amino acid residue
- $AA_9$  is a neutral/polar/small amino acid residue; and
- $Z_2$  is hydroxy-, carboxy-, non-amino acids such as agmatine, or at least one amino acid residue, including carboxamide or alkylamide forms thereof.

2. The method of Claim 1 wherein:

- $AA_1$  is tryptophan or n-formyl-tryptophan;
- $AA_2$  is serine, threonine or aspartic acid;
- $AA_3$  is proline, glutamic acid, serine or isoleucine;
- $AA_4$  is cysteine;
- $AA_5$  is serine or asparagine;
- $AA_6$  is valine or threonine;
- $AA_7$  is threonine or serine;
- $AA_8$  is cysteine;
- $AA_9$  is glycine or serine, including carboxamide forms thereof

3. The method of Claim 2 wherein the polypeptide compound is selected from the group consisting of:

W-S-P-C-S-V-T-C-G (SEQ ID No. 1),  
 FW-S-P-C-S-V-T-C-G-NH<sub>2</sub>,  
 5 W-S-P-C-S-V-T-C-G-NH<sub>2</sub>,  
 W-D-I-C-S-V-T-C-G (SEQ ID No. 2)  
 W-S-S-C-S-V-T-C-G (SEQ ID No. 3),  
 10 W-T-S-C-S-T-S-C-G (SEQ ID No. 4),  
 W-S-P-W-S-E-W-T-S-C-S-T-S-C-G-N-G-I-Q-Q-R-G-R (SEQ ID No. 5),  
 W-S-H-W-S-P-W-S-S-C-S-V-T-C-G-D-G-V-I-T-R-I-R (SEQ ID No. 6),  
 15 W-G-P-W-S-P-W-D-I-C-S-V-T-C-G-G-G-V-Q-K-R-S-R (SEQ ID No. 7),  
 W-S-P-C-S-V-T-C-S (SEQ ID No. 8),  
 W-S-Q-C-S-V-T-C-G (SEQ ID No. 9),  
 W-S-Q-C-N-V-T-C-G (SEQ ID No. 10),  
 20 W-T-P-C-S-V-T-C-G (SEQ ID No. 11), and  
 D-G-G-W-S-H-W-S-P-W-S-S-C-S-V-T-C-G-D-G-V-I-T-R-  
 I-R-L-C-N-S-P-S-P-Q-M-N-G-K-P-C-E-G-E-A-R-E-T-K-A-  
 25 C-K-K-D-A-C-P-I-N-G-G (SEQ ID No. 12)  
 W-D-E-C-R-Q-T-C-G-A-S (SEQ ID No. 13),  
 W-S-A-C-S-L-G-C-D (SEQ ID No. 14),  
 30 W-D-I-C-S-V-T-C-G-NH<sub>2</sub>,  
 W-S-S-C-S-V-T-C-G-NH<sub>2</sub>, and  
 W-T-S-C-S-T-S-C-G-NH<sub>2</sub>

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4. The method of Claim 1 which inhibits tumor cell metastasis.
5. The method of Claim 1 which promotes or inhibits platelet aggregation.
- 40 6. The method of Claim 1 which promotes or inhibits angiogenic activity.
7. The method of claim 1 which promotes or inhibits thrombotic activity.
- 45 8. The method of Claim 1 which promotes wound healing.
9. The method of Claim 1 which inhibits atherosclerosis.
10. The method of Claim 1 which promotes or inhibits thrombolytic activity.
- 50 11. The method of Claim 1 which promotes or inhibits immune modulation.
12. The method of Claim 1 which promotes or inhibits cell adhesion.
- 65 13. The method of Claim 1 wherein said polypeptide compound is admixed with at least one pharmaceutically acceptable carrier prior to being administered.
14. A composition useful to promote or inhibit thrombospondin-like activity comprising a pharmaceutically

acceptable carrier together with an effective amount of a polypeptide compound comprising the formula:



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wherein:

- $Z_1$  is hydrogen, amino, acetyl, or at least one amino acid residue or the desamino forms thereof;
- $AA_1$  is a neutral/nonpolar/large/cyclic amino acid residue;
- 10  $AA_2$  is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;
- $AA_3$  is a neutral/nonpolar/large/cyclic or neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- $AA_4$  is a neutral/polar/small amino acid residue;
- $AA_5$  is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;
- 15  $AA_6$  is a neutral/nonpolar/large/non-cyclic/neutral/polar/large/non-cyclic amino acid residue;
- $AA_7$  is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- $AA_8$  is a neutral/polar/small amino acid residue
- $AA_9$  is a neutral/polar/small amino acid residue; and
- 20  $Z_2$  is hydroxyl, carboxyl, non-amino acids such as agmatine, or at least one amino acid residue, including carboxamide or alkylamide forms thereof.

15. Antisera and antibodies which are capable of recognizing and specifically binding to an immunoreactive polypeptide comprising the formula:



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wherein:

- $Z_1$  is hydrogen, amino, acetyl, or at least one amino acid residue or the desamino forms thereof;
- 30  $AA_1$  is a neutral/nonpolar/large/cyclic amino acid residue;
- $AA_2$  is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;
- $AA_3$  is a neutral/nonpolar/large/cyclic or neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- $AA_4$  is a neutral/polar/small amino acid residue;
- 35  $AA_5$  is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;
- $AA_6$  is a neutral/nonpolar/large/non-cyclic/neutral/polar/large/non-cyclic amino acid residue;
- $AA_7$  is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- $AA_8$  is a neutral/polar/small amino acid residue
- $AA_9$  is a neutral/polar/small amino acid residue; and
- 40  $Z_2$  is hydroxyl, carboxyl, non-amino acids such as agmatine, or at least one amino acid residue, including carboxamide or alkylamide forms thereof.

16. A method for facilitating the delivery of compounds for diagnostic or therapeutic purposes comprising a polypeptide compound comprising the formula:

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wherein:

- $Z_1$  is hydrogen, amino, acetyl, or at least one amino acid residue or the desamino forms thereof;
- 50  $AA_1$  is a neutral/nonpolar/large/cyclic amino acid residue;
- $AA_2$  is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;
- $AA_3$  is a neutral/nonpolar/large/cyclic or neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;
- 55  $AA_4$  is a neutral/polar/small amino acid residue;
- $AA_5$  is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;
- $AA_6$  is a neutral/nonpolar/large/non-cyclic/neutral/polar/large/non-cyclic amino acid residue;
- $AA_7$  is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;

AA<sub>8</sub> is a neutral/polar/small amino acid residue  
 AA<sub>9</sub> is a neutral/polar/small amino acid residue; and  
 Z<sub>2</sub> is hydroxyl, carboxyl, non-amino acids such as agmatine, or at least one amino acid residue, including carboxamide or alkylamide forms thereof.

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17. A biomedical device for promoting the attachment of cells which incorporates a polypeptide compound comprising the formula:

Z<sub>1</sub>-AA<sub>1</sub>-AA<sub>2</sub>-AA<sub>3</sub>-AA<sub>4</sub>-AA<sub>5</sub>-AA<sub>6</sub>-AA<sub>7</sub>-AA<sub>8</sub>-AA<sub>9</sub>-Z<sub>2</sub>

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wherein:

Z<sub>1</sub> is hydrogen, amino, acetyl, or at least one amino acid residue or the desamino forms thereof;

AA<sub>1</sub> is a neutral/nonpolar/large/cyclic amino acid residue;

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AA<sub>2</sub> is a neutral/polar/small or neutral/polar/large/non-cyclic or acidic amino acid residue;

AA<sub>3</sub> is a neutral/nonpolar/large/cyclic or neutral/nonpolar/large/non-cyclic or neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;

AA<sub>4</sub> is a neutral/polar/small amino acid residue;

AA<sub>5</sub> is a neutral/polar/small or neutral/polar/large/non-cyclic amino acid residue;

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AA<sub>6</sub> is a neutral/nonpolar/large/non-cyclic/neutral/polar/large/non-cyclic amino acid residue;

AA<sub>7</sub> is a neutral/polar/large/non-cyclic or neutral/polar/small amino acid residue;

AA<sub>8</sub> is a neutral/polar/small amino acid residue

AA<sub>9</sub> is a neutral/polar/small amino acid residue; and

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Z<sub>2</sub> is hydroxyl, carboxyl, non-amino acids such as agmatine, or at least one amino acid residue, including carboxamide or alkylamide forms thereof.

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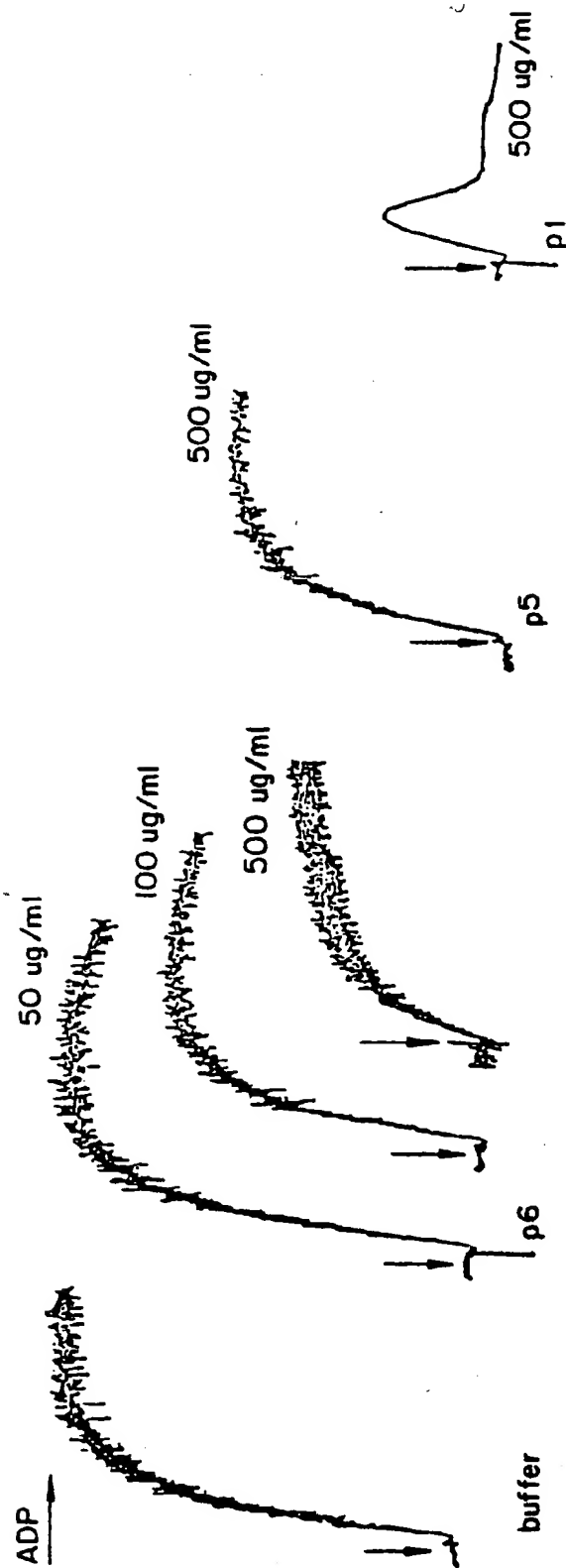


FIG. 1



European Patent  
Office

**PARTIAL EUROPEAN SEARCH REPORT**  
which under Rule 45 of the European Patent Convention  
shall be considered, for the purposes of subsequent  
proceedings, as the European search report

Application number

EP 91 10 1908

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
P,X	WO-A-90 01 496 (RESEARCH EXPLOITATION LTD)  * Whole document and particul. claim 5 *	14-15	C 07 K 7/00 A 61 K 37/02 C 07 K 15/00
D,A	CHEMICAL ABSTRACTS, vol. 106, 1987, page 141, abstract no. 62011v, Columbus, Ohio, US; J. LAWLER et al.: "The structure of human thrombospondin, an adhesive glycoprotein with multiple calcium-binding sites and homologies with several different proteins", & J. CELL. BIOL., 1986, 103(5, Pt. 1), 1635-48  * Abstract *	14-15	TECHNICAL FIELDS SEARCHED (Int. Cl.4)  C 07 K A 61 K
INCOMPLETE SEARCH			
<p>The Search Division considers that the present European patent application does not comply with the provisions of the European Patent Convention to such an extent that it is not possible to carry out a meaningful search into the state of the art on the basis of some of the claims.</p> <p>Claims searched completely: 14-17 Claims searched incompletely: Claims not searched: 1-13 Reason for the limitation of the search:</p> <p>Method for treatment of the human or animal body by surgery or therapy. (See art. 52(4) of the European Patent Convention)</p>			
Place of search THE HAGUE		Date of completion of the search 03-06-1991	Examiner RAJIC
<p>CATEGORY OF CITED DOCUMENTS</p> <p>X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document</p> <p>T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons &amp; : member of the same patent family, corresponding document</p>			